

## Stereoselective Synthesis of 5-Methylphosphono-D-Arabino Hydroximolactone, Inhibitor of Glucosamine-6-Phosphate Synthase and Phosphoglucose Isomerase.

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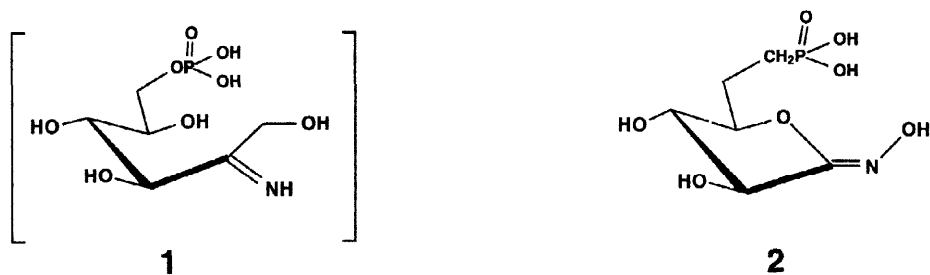
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**Abstract :** Compound **2**, synthesized from D-arabinose in 12 steps with an overall 4% yield, is a competitive inhibitor vs fructose-6P for both phosphoglucose isomerase and glucosamine-6P synthase.

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The reaction performed by glucosamine-6P synthase (L-glutamine: D-fructose-6-phosphate amidotransferase, GlmS), *i.e.* conversion of fructose-6P into glucosamine-6P, is believed to proceed through the formation of intermediate **1**, a Schiff base between the keto group of the sugar and the ammonia generated from the glutamine amide function<sup>1</sup>. This intermediate is then processed by the keto/aldose isomerase activity of GlmS to give glucosamine-6P. In the course of our search for specific inhibitors of this enzyme involved in microbial cell wall biosynthesis, the phosphonate **2** was considered to mimick reasonably well the putative intermediate **1** without presenting the sensitivity of the phosphate bond towards hydrolysis.

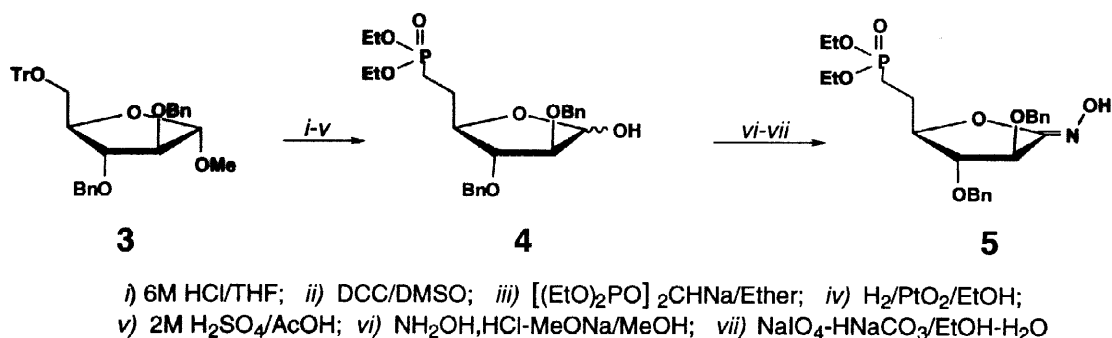


Scheme 1

We now describe the synthesis of **2** and its evaluation *in vitro* against *Escherichia coli* GlmS and commercially available rabbit muscle phosphoglucose isomerase (PGI), another member of the keto/aldose isomerase family using fructose-6P as a substrate.

Fully protected D-arabinose **3** (scheme 2), obtained according to literature (Mp 80–82°C, lit<sup>2</sup> 80–81°C), was detritylated in 93% yield with 6M HCl in THF<sup>3</sup>. The resulting primary alcohol was oxidized under Moffatt conditions<sup>4</sup> to afford the corresponding aldehyde which was used without further purification. The methyl phosphonate was isolated in 51% yield from the aldehyde following Horner-Emmons condensation with the tetraethyl methylenediphosphonate carbanion<sup>5</sup> (sodium hydride-ether) and hydrogenation over PtO<sub>2</sub>. Hydrolysis with sulfuric acid (2M in AcOH, 60°C) afforded **4** in 73% yield as a mixture of non separable anomers. **4** was readily oxidized into the corresponding lactone unwilling, however, to give a stable adduct upon condensation with hydroxylamine.

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Scheme 2

Inverting the sequence of these two steps, *i.e.* reaction of **4** with hydroxylamine and oxidation by sodium metaperiodate (1.1 equivalents) in the presence of sodium bicarbonate<sup>6</sup>, gave the protected hydroximolactone **5** in 34 % yield for the two steps.

Phosphonate ester hydrolysis was accomplished using neat bromotrimethylsilane<sup>7</sup> followed by anhydrous triethylamine quenching of the reaction mixture. Hydrogenolysis of the benzyl groups was finally performed (MeOH, 1 eq. triethylamine) with a large excess of Pd/C (1:1 by weight) and hydrogen gas, to give the hydroximolactone **2** in 50% yield as a monotriethyl ammonium salt after lyophilization<sup>8</sup>.

When tested against purified *Escherichia coli* GlmS<sup>9</sup> or commercially available rabbit PGI, **2** behaved as a competitive inhibitor vs fructose-6P with respective  $K_i$  of 0.4 mM and 32  $\mu\text{M}$ . These values reflect an affinity of **2** for the enzyme active site similar to that of the substrate ( $K_m/K_i = 1$  and 3.1 respectively). Since PGI is known to recognize the cyclic form of fructose-6P<sup>10</sup>, it can be concluded that GlmS might operate in the same way before undergoing Schiff base formation between fructose-6P and lysine 603<sup>1</sup>.

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- Selected analytical data for **2**:  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  1.4-2.1 (4H, m, H5-H6), 4.05 (1H, t, H3), 4.3 (1H, m, H4), 4.85 (1H, d, H2).  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  25.8-27.6 (C6,  $J_{\text{C-P}} = 131$  Hz), 29.7 (C5), 76.5 (C2), 80.2 (C3), 88.3 (C4), 162 (C1).  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ , ref  $\text{K}_2\text{HPO}_4$ )  $\delta$  22.9. MS (negative FAB, glycerol) 240 (M-H)<sup>-</sup>, 183.
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